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# Direct Lipase Catalyzed Lipophilization of Chlorogenic Acid in Supercritical Carbon Dioxide

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## ABSTRACT

The enzymatic lipophilization of natural antioxidants is a process of enormous pharmaceutical interest; the direct esterification of such natural antioxidants is considered a major challenge due to the heterogeneity of these reactants. In this study, a Taguchi approach was employed to optimize the lipophilization of chlorogenic acid using immobilized *Candida antarctica* lipase. The effects of reaction temperature (35-55 °C), pressure (150-250 bar), enzyme amount (10-30 mg/ml), and reaction time (10-25 hours) were investigated. The maximum conversion reached 63 % in 25 hours at 150 bar, 55 °C, 10 % *t*-butanol (v/v), and 20 mg/ml of lipase. The SC CO<sub>2</sub> selectivity towards the esterified product was the working principle of this study, by which minimized interphase transport limitations and enhanced mass-transfer phenomena substantially improved the reaction kinetics. This investigation offers an alternative towards the functionalization of natural antioxidants which harmonizes with the use of green technologies.

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Abstract requires 140-170 words  
and only can have one

Keywords need in alphabetic order.

**Key words:** *Candida antarctica* lipase, Chlorogenic acid, lipophilization, supercritical

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## Introduction

Chlorogenic acid (5-caffeoyl-quinic acid, 5-CGA) has been associated to a broad range of bioactivities (Bonita *et al.*, 2007; Clifford, 2000; Zheng *et al.*, 2008), while its consumption from coffee water extracts determined to be safe (Watanabe *et al.*, 2006). Biological properties of 5-CGA are primary attributed to its capacity to donate hydrogen atoms of the phenolic ring to free radicals, through various processes. It has been proposed that such bioactivities of natural phenolics can be enhanced by increasing their amphiphilic structured phenolic would incorporate both emulsifying and antioxidant properties (Figuroa-Espinoza and Villeneuve, 2005; Jayaprakasam *et al.*, 2006; Sabally *et al.*, 2006; Vosmann, Weitkamp, and Weber, 2006; Weitkamp, Vosmann, and Weber, 2006).

## Materials and Methods

### Materials

Immobilized lipase from *Candida antarctica* (lipase B, Novozym 435) was provided by Novo Nordisk. Chlorogenic acid (5-CGA), 2-methyl-2-propanol (*t*-butanol), 1-heptanol, 1-pentanol, and geraniol were obtained from Sigma-Aldrich. All solvents used were HPLC grade and from Merck.

### Lipase Catalyzed Esterification in SC CO<sub>2</sub>

A batch-operated stirred-system was specially designed and built in our laboratory to carry out the enzymatic reaction. A cell volume of 50 mL is described in Figure 1.

## Results and Discussion

In this work, the possibility of performing a lipase-catalyzed esterification of 5-CGA and 1-heptanol in SC CO<sub>2</sub>, while simultaneously extracting the formed compound, was demonstrated.

The biotransformation of 5-CGA via esterification with 1-heptanol was used to incorporate the corresponding seven-carbon aliphatic chain, and as a result modify its polarity. As shown by HPLC analysis of samples withdrawn at specific intervals from the reaction mixture (Figure 2), lipophilization resulted in a significant increase of the acid original retention time (from 24 to 38 min) and Log P values (from -0.75 to 2).

Indent two Chinese characters (4 English characters) at the beginning of every paragraph.

*et al.*, : in citations, only with four or more authors use *et al.*  
\* *et al.* should be italic

Citation format:

Only put the last name of author in the parentheses.

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The resulting modified lipophilicity consequently enhanced the selectivity of the reaction medium towards the formed ester.

### **Conclusions**

The Taguchi experimental design provided valuable insights into the lipase-catalyzed esterification of 5-CGA in SC CO<sub>2</sub>.

Immobilized lipase B from *Candida antarctica* (20 mg/ml) was used to successfully lipophilize 5-CGA by esterification with 1-heptanol in SCO<sub>2</sub>/*t*-butanol (10 % v/v) at 150 bar, and 55 °C. Under these conditions, reaction rates approached 2.32 μM ester/ g lipase per minute with conversions of 63 %.

The Reference format: Place "References,"  
alphabetically by authors.

## References

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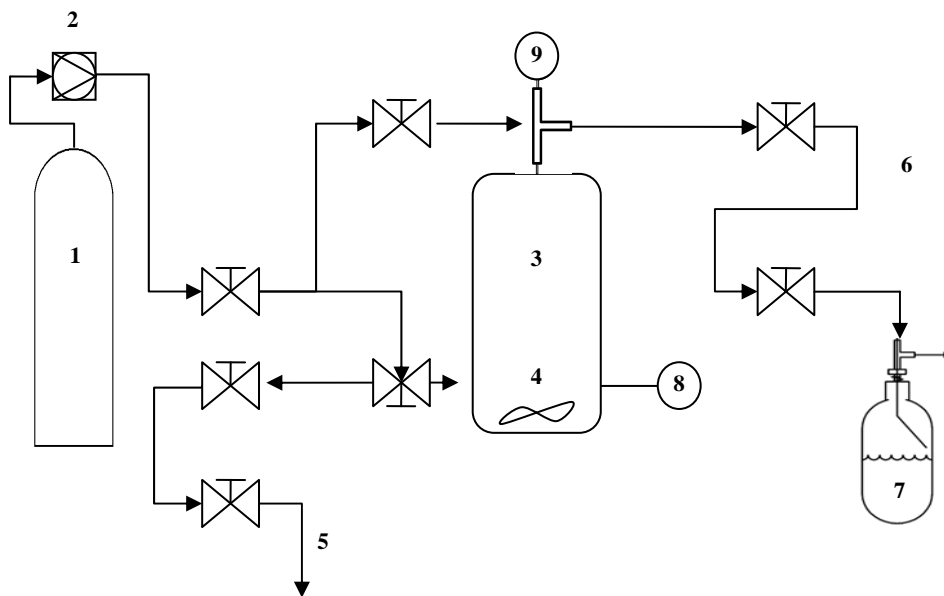
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**Table 1. Parameters and Levels Used in This Experiment**

**Experimental Design**

Parameter	Level		
	1	2	3
Temperature (°C)	35	45	55
Pressure (bar)	150	200	250
LiOH (%)	2	5	10
Enzyme (mg/ml)	10	20	30

Present tables with the minimum use of horizontal rules (usually three are sufficient) and avoiding vertical rules except in matrices.



**Figure 1. Scheme of the Experimental Batch-Stirred Apparatus for Synthesis Under High Pressure: (1) CO<sub>2</sub> Tank, (2) High Pressure Pump, (3) Reactor, (4) Stirrer, (5) SC CO<sub>2</sub> Sampling Loop (From Bottom of the Reactor), (6) SC CO<sub>2</sub> Sampling Loop (From Top of the Reactor), (7) Collector, (8) Temperature Controller, (9) Pressure Gauge**

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